DESCRIPTION

HEPATOCELLULAR CARCINOMA-ASSOCIATED GENE

TECHNICAL FIELD

The present invention relates to a gene associated with hepatocellular carcinoma, and particularly to a gene associated with the recurrence of hepatocellular carcinoma.

BACKGROUND ART

Almost all types of hepatocellular carcinomas are developed from chronic hepatitis caused by viral hepatitis. The causal viruses thereof are hepatitis C virus and hepatitis B virus. If a patient is persistently infected with either hepatitis C virus or hepatitis B virus, there are no therapeutic methods therefor. The patient does nothing but only facing a fear of developing liver cirrhosis or hepatocellular carcinoma. Interferon has been used as an agent for treating hepatitis. However, effective examples are only 30%, and thus this is not necessarily a sufficient therapeutic agent. Under the present circumstances, there are almost no effective examples, in particular, for chronic hepatitis. Nevertheless, even if such viruses cannot be eliminated, if progression of pathologic conditions can be suppressed, it leads to prevention of liver cirrhosis or hepatocellular carcinoma. Thus, it is considered important to clarify the factor of developing pathologic conditions at a molecular level.

If once hepatocellular carcinoma has been developed, even if a surgical radical operation is made, the recurrence of cancer in the remaining liver appears at a high frequency. The survival rate obtained 5 years after the operation of liver cancer is 51% on a national accumulation base. It has been reported that such recurrence appears at approximately 25% of cases 1 year after hepatectomy, at 50% thereof 2 years after hepatectomy, and at 80% thereof 5 years after hepatectomy. Hence, it cannot be said that remaining liver tissues are normal liver tissues, but it is considered that a bud of the

recurrence of hepatocellular carcinoma has already existed. At present, it has been reported that recurrence risk factors include the maximum diameter of a tumor, the number of tumors, tumor embolus of portal vein, a preoperative AFP value, intrahepatic metastasis, the presence or absence of liver cirrhosis, etc. However, in order to develop a method for predicting and preventing the recurrence of hepatocellular carcinoma, it is necessary to find at a molecular level a factor of determining the presence or absence of recurrence, which is associated with such risk factors. Such a factor obtained at a molecular level is considered to be a factor, which is associated not only with recurrence but also with the development of hepatocellular carcinoma or progression of pathologic conditions. In recent years, as a result of gene expression analysis using a DNA microarray, it has become possible to classify more in detail such pathologic conditions based on the difference in the expression patterns of genes as a whole. To date, histological or immunological means have been mainly used for classification of cancers. However, cancers classified into the same type have different clinical courses and therapeutic effects depending on individual cases. If there were a means for classifying such cancers more in detail, it would become possible to offer treatment depending on individual cases. It is considered that the gene expression analysis using a DNA microarray constitutes a powerful method for knowing the prognosis of such cancers.

To date, the DNA microarray analysis has clarified the following points associated with hepatocellular carcinoma:

(i) the types of genes, the expressions of which are different between a tumor tissue and a nontumor tissue (Shirota Y, Kaneko S, Honda M, et al. Identification of differentially expressed gene in hepatocellular carcinoma with cDNA microarrays. Hepatology 2001; 33: 832-840, Xu X, Huang J, Xu Z, et al. Insight into hepatocellular carcinogenesis at transcriptome level by comparing gene expression profiles of hepatocellular carcinoma with those of corresponding noncancerous liver. Proc. Nat. Acad. Sci. USA. 2001; 98: 15089-15094);

(ii) in terms of the differentiation degree of cancer tissues, the types of genes, the

expressions of which are different (Shirota Y, Kaneko S, Honda M, et al. Identification of differentially expressed gene in hepatocellular carcinoma with cDNA microarrays. Hepatology 2001; 33: 832-840, Okabe H, Satoh S, Kato T, et al. Genome-wide analysis of gene expression in human hepatocellular carcinomas using cDNA microarray: Identification of genes involved in viral carcinogenesis and tumor progression. Cancer res. 2001 ; 61 : 2129- 2137);

(iii) the types of genes, the expressions of which are different between hepatocellular carcinoma derived from hepatitis B and hepatocellular carcinoma derived from hepatitis C (Okabe H, Satoh S, Kato T, et al. Genome-wide analysis of gene expression in human hepatocellular carcinomas using cDNA microarray: Identification of genes involved in viral carcinogenesis and tumor progression. Cancer res. 2001; 61: 2129- 2137);

(iv) the types of genes, the expressions of which are different depending on the presence or absence of vascular invasion of hepatocellular carcinoma (Okabe H, Satoh S, Kato T, et al. Genome-wide analysis of gene expression in human hepatocellular carcinomas using cDNA microarray : Identification of genes involved in viral carcinogenesis and tumor progression. Cancer res. 2001; 61: 2129- 2137); and

(v) the type of a change in gene expression observed among intrahepatic metastatic cancers, as a result of the clonal analysis of multinodular hepatocellular carcinoma (Cheung S, Chen X, Guan X, et al. Identify metastasis-associated gene in hepatocellular carcinoma through clonality delineation for multinodular tumor. Cancer res. 2002; 62: 4711- 4721).

However, with regard to genes associated with recurrence, only the analysis of Iizuka et al. on cancer tissues has existed (Iizuka N, Oka M, Yamada-Okabe H, et al. Oligonucleotide microarray for prediction of early intrahepatic recurrence of hepatocellular carcinoma after curative resection. Lancet 2003; 361: 923-929). The analysis of nontumor liver tissues, which reflects the remaining liver tissues, has not yet been achieved.

DISCLOSURE OF THE INVENTION

It is an object of the present invention to provide a gene associated with hepatocellular carcinoma, and particularly, a gene, which predicts the recurrence of the cancer.

As a result of intensive studies directed towards achieving the aforementioned object, the present inventor has studied the profile of gene expression based on a case where hepatocellular carcinoma has recurred and a case where hepatocellular carcinoma has not recurred, and has succeeded in identification of a gene associated with hepatocellular carcinoma, thereby completing the present invention.

That is to say, the present invention has the following features:

(1) A method for evaluating cancer, which comprises the following steps of:

(a) collecting total RNA from an analyte;

(b) measuring the expression level of at least one gene selected from among the genes shown in Tables 1 to 8; and

(c) evaluating cancer using the measurement result as an indicator.

In the present invention, from among the genes shown in Tables 1 to 8, at least one gene selected from the group consisting of the PSMB8 gene, the RALGDS gene, the GBP1 gene, the RPS14 gene, the CXCL9 gene, the DKFZp564F212 gene, the CYP1B1 gene, the TNFSF10 gene, the NR0B2 gene, the MAFB gene, the BF530535 gene, the MRPL24 gene, the QPRT gene, the VNN1 gene, and the IRS2 gene, can be used, for example. Otherwise, from among the genes shown in Tables 1 to 8, at least one gene selected from the group consisting of the PZP gene, the MAP3K5 gene, the TNFSF14 gene, the LMNA gene, the CYP1A1 gene, and the IGFBP3 gene, can be used, for example.

In addition, when such measurement is carried out using GAPDH as an internal standard gene, from among the genes shown in Tables 1 to 8, each gene contained in a gene set consisting of the VNN1 gene and the MRPL24 gene, or a gene set consisting of the PRODH gene, the LMNA gene, and the MAP3K12 gene, can be used.

Moreover, when such measurement is carried out using 18S rRNA as an internal standard gene, from among the genes shown in Tables 1 to 8, each gene contained in a gene set consisting of the VNN1 gene, the CXCL9 gene, the GBP1 gene, and the RALGDS gene, or a gene set consisting of the LMNA gene, the LTBP2 gene, the COL1A2 gene, and the PZP gene, can be used.

The above evaluation of cancer involves prediction of the presence or absence of metastasis or recurrence. Further, an example of such cancer is hepatocellular carcinoma.

The expression level of a gene can be measured by amplifying the gene, using at least one set of primers consisting of the nucleotide sequences shown in SEQ ID NOS: 2n-1 and 2n (wherein n represents an integer between 1 and 114). Otherwise, the expression level of a gene can be measured by amplifying the gene, using a set of primers for amplifying each gene contained in at least one gene set selected from the group consisting of a gene set consisting of the VNN1 gene and the MRPL24 gene, a gene set consisting of the PRODH gene, the LMNA gene, and the MAP3K12 gene, a gene set consisting of the VNN1 gene, the CXCL9 gene, the GBP1 gene, and the RALGDS gene, and a gene set consisting of the LMNA gene, the LTBP2 gene, the COL1A2 gene, and the PZP gene.

(2) A primer set, which comprises at least one set of primers consisting of the nucleotide sequences shown in SEQ ID NOS: 2n-1 and 2n (wherein n represents an integer between 1 and 114).

(3) A primer set, which comprises a set of primers for amplifying each gene contained in at least one gene set selected from the group consisting of a gene set consisting of the VNN1 gene and the MRPL24 gene, a gene set consisting of the PRODH gene, the LMNA gene, and the MAP3K12 gene, a gene set consisting of the VNN1 gene, the CXCL9 gene, the GBP1 gene, and the RALGDS gene, and a gene set consisting of the LMNA gene, the LTBP2 gene, the COL1A2 gene, and the PZP gene.

(4) A kit for evaluating cancer, which comprises any gene shown in Tables 1 to 8.

An example of the aforementioned gene is at least one gene selected from the group consisting of the RALGDS gene, the GBP1 gene, the DKFZp564F212 gene, the TNFSF10 gene, and the QPRT gene.

Moreover, another example of the aforementioned gene is each gene contained in at least one gene set selected from the group consisting of a gene set consisting of the VNN1 gene and the MRPL24 gene, a gene set consisting of the PRODH gene, the LMNA gene, and the MAP3K12 gene, a gene set consisting of the VNN1 gene, the CXCL9 gene, the GBP1 gene, and the RALGDS gene, and a gene set consisting of the LMNA gene, the LTBP2 gene, the COL1A2 gene, and the PZP gene.

Furthermore, the kit of the present invention may comprise the aforementioned primer set.

The present invention provides a gene useful for predicting the recurrence of hepatocellular carcinoma. Cancer can be evaluated by analyzing the increased expression state of such a gene. In particular, using the gene of the present invention, the recurrence of hepatocellular carcinoma can be predicted, and the obtained prediction information is useful for the subsequent therapeutic strategy. Moreover, the use of such a gene and a gene product enables the development of a treatment method for preventing recurrence.

BRIEF DESCRIPTION OF THE DRAWING

Figure 1 is a view showing the phylogenetic tree of samples obtained from the entire gene expression profile. Genes are rearranged based on the similarity in expression manner among samples, and further, samples are rearranged based on the similarity in the expression manner of the entire genes. Thus, the genetic affiliation is expressed in the form of a phylogenetic tree.

BEST MODE FOR CARRYING OUT THE INVENTION

The present invention will be described in detail below.

The present invention is characterized in that the follow-up clinical data collected for a long period of time after the resection of hepatocellular carcinoma are divided into a poor prognosis case group (for example, a case group wherein the cancer recurs within 1 year, leading to death within 2 years) and into a good prognosis case group (for example, a case group wherein the cancer does not recur for 4 or more years), and is characterized in that a gene causing poor prognosis or a gene causing good prognosis (for example, a gene associated with promotion of the recurrence and a gene associated with suppression of the recurrence) is identified based on the characteristics of a gene group, which is expressed in the excised liver tissues. The present invention relates to classification of causal viruses into type B hepatocellular carcinoma cases and into type C hepatocellular carcinoma cases based on clinical data, and identification of a gene having a prognostic correlation from each of the tissues of a nontumor tissue and the tissues of a tumor tissue.

The gene of the present invention is obtained by analyzing the correlation between tissues actually collected from a patient and a pathologic condition thereof, and thereby clarifying the type of a case, a pathologic condition, and a gene, which are used to clarify the correlation between a gene and a pathologic condition.

1. Classification of test samples

The postoperative course is observed after an operation to resect liver cancer, and test samples are classified into an early recurrence group and into a late recurrence group.

The term "early recurrence group" is used to mean a case group wherein the cancer recurs within a certain period of time after resection, thereafter leading to death. A recurrence period is not particularly limited. For example, it is 1 year or shorter, or 2 years or shorter. A survival time is not particularly limited either. For example, it is 1 year or shorter, 2 years or shorter, or 3 years or shorter, after recurrence. The term "late

recurrence group" is used to mean a case group wherein the cancer does not recur for a certain period of time after resection (for example, 3 years or longer, and preferably 4 years or longer).

In reality, 51 cases, which were subjected to an operation to resect hepatocellular carcinoma at stages I and II, were used as targets. The 51 cases contain 16 cases of type B hepatocellular carcinoma and 35 cases of type C hepatocellular carcinoma. Based on the follow-up clinical data of such cases, 2 cases were selected from the type B hepatocellular carcinoma and 3 cases were selected from the type C hepatocellular carcinoma, and these cases were classified into an early recurrence group. On the other hand, 2 cases selected from the type B hepatocellular carcinoma and 3 cases were selected from the type C hepatocellular carcinoma, and these cases were classified into a late recurrence group. With regard to the RNA portions of the nontumor tissues and tumor tissues of such 10 cases, the following expression profile analysis was carried out.

2. Gene analysis

Total RNA is extracted from each type of the liver tissues of the classified groups, and gene expression profiles are then compared between the groups using a microarray. Such total RNA can be extracted using a commercially available reagent (for example, TRIzol). For detection of an expression profile, Microarray (Affymetrix) is used, for example.

Moreover, the present invention enables the analysis of a gene, which changes expression in the tissues of a nontumor tissue as well as in the tissue of a tumor tissue. The term "nontumor tissue" is used herein to mean liver tissues involved in a resection of hepatocellular carcinoma, which do not contain cancer cells. However, such a "nontumor tissue" does not necessarily mean normal liver tissues, but it also includes tissues affected by chronic hepatitis (hepatitis B or hepatitis C) or liver cirrhosis. For example, a gene up-regulated in a nontumor tissue in a late recurrence group including type B hepatocellular carcinoma cases or type C hepatocellular carcinoma cases, wherein

almost all tissues are such affected tissues, can be used as an analysis target. In the case of such tissues affected by chronic hepatitis or liver cirrhosis, a necrotic inflammatory reaction, regenerating nodules, fibrosis attended with decidual liver cells, or the like are observed. Among such cells, there are cells, which can be potential cells causing the development of hepatocellular carcinoma. Accordingly, it is considered that gene expression relevant to prognosis exists in the nontumor tissue. Thus, prognosis (for example, recurrence) can be predicted using such gene expression as an indicator (for example, by analyzing changes in such gene expression).

A gene used for evaluation of cancer is identified based on the correlation of changes in gene expression with phenotype (recurrence, early progression, etc.). The term "evaluation of cancer" is used to mean evaluation regarding the pathologic conditions of cancer or the stage of cancer progression. Such evaluation of cancer includes prediction of the presence or absence of metastasis or recurrence.

The present invention provides an up-regulated gene or a down-regulated gene in terms of recurrence. The term "recurrence" is used to mean that a lesion, which is considered to be a new carcinoma, appears in the liver, after a treatment for a primary lesion has been determined to complete.

3. Evaluation of gene

Using disease model cells or animals, the identified gene is evaluated in terms of availability as a factor of suppressing the development of pathologic conditions. Namely, (1) the remaining cases of hepatocellular carcinoma, the prognosis of which has been known, are subjected to quantitative analysis of gene expression, and the correlation with the prognosis is studied. (2) The gene is transferred into a hepatocellular carcinoma-cultured cell line, and it is allowed to express therein. Thereafter, the cell growth and a change in malignancy are evaluated based on ability to form colonies in a soft agar plate or ability to form tumors in nude mice. (3) Using a cultured hepatic cell line established from a patient with chronic hepatitis, the gene is

transferred into the cells, and it is allowed to express therein. Thereafter, the cell growth and malignant transformation are evaluated by the same method as that described in (2) above. (4) The gene is transferred into the liver of a hepatocellular carcinoma development-model animal, and it is allowed to express therein. Thereafter, the course up to the development of liver cancer is evaluated.

In (1) above, the quantitative analysis of gene expression is carried out by real-time PCR, for example. That is to say, a commercially available reverse transcriptase is used for the total RNA as produced above, so as to synthesize cDNA. As a PCR reagent, a commercially available reagent can be used. Moreover, PCR may be carried out in accordance with commercially available protocols. For example, preliminary heating is carried out at 95°C for 10 minutes, and thereafter, a cycle consisting of 95°C for 15 seconds and 60°C (or 65°C) for 60 seconds, is repeated 40 times. Examples of an internal standard gene used herein as a target may include housekeeping genes such as glyceraldehyde 3-phosphatase dehydrogenase (GAPDH), 18S ribosomal RNA (18S rRNA), β-Actin, cyclophilin A, HPRT1 (hypoxanthine phosphoribosyltransferase 1), B2M (beta-2 microglobulin), ribosomal protein L13a, or ribosomal protein L4. Persons skilled in the art can appropriately select such an internal standard gene. As an analysis method, absolute quantitative analysis or relative quantitative analysis of an expression level is adopted. The absolute quantitative analysis is preferable. Herein, absolute quantification of an expression level is obtained by determining a threshold line on which a calibration curve becomes optimum and then obtaining the number of threshold PCR cycles and a threshold cycle value (Ct) of each sample. On the other hand, a relative expression level is expressed with a Δ Ct value obtained by subtracting the Ct value of an internal standard gene (for example, GAPDH) from the Ct value of a target gene. Values obtained using the formula ($2^{(-\Delta Ct)}$) can be used for evaluation of a linear expression level.

When a calibration curve is produced, values obtained by subjecting standard samples to serial dilution and simultaneous measurement (the samples are placed in a

single plate and simultaneously measured, using a single reaction solution) may be used.

When an absolute expression level can be obtained relative to a calibration curve, the absolute expression level of a target gene and that of an internal standard gene are obtained, and the ratio of the target gene expression level/the internal standard gene expression level is calculated for each sample, so as to use it for evaluation.

Genes are selected from the results of the microarray of a late recurrence group and that of an early recurrence group. Thereafter, among genes, regarding which the results of real-time PCR obtained by the aforementioned method correspond with the results of the microarray, those exhibiting a correlation with a recurrence period can be identified as up-regulated genes of nontumor tissue, for example.

As described above, as genes identified as an up-regulated gene, various genes can be selected depending on experimental conditions applied during the identification, such as an internal standard gene, a primer sequence, or an annealing temperature which are used. Also, using various types of statistical methods (for example, Mann-Whitney U test), a gene correlating to a recurrence period can be selected.

The full-length sequence of the gene of the present invention can be obtained as follows. That is to say, it is searched through DNA database, and it can be obtained as known sequence information. Otherwise, the above full-length sequence is isolated from human liver cDNA library by hybridization screening.

In the present invention, genes up-regulated in cases where the cancer has not recurred at an early date (late recurrence) include those shown in Tables 1 to 4. On the other hand, genes up-regulated in cases where the cancer has recurred at an early date include those shown in Tables 5 to 8.

Table 1: Genes (24) up-regulated in a nontumor tissue in a late recurrence group of type B hepatocellular carcinoma cases

Table 2: Genes (10) up-regulated in a nontumor tissue in a late recurrence group of type C hepatocellular carcinoma cases

Table 3: Genes (137) up-regulated in a tumor tissue in a late recurrence group of type B

hepatocellular carcinoma cases

Table 4: Genes (104) up-regulated in a tumor tissue in a late recurrence group of type C hepatocellular carcinoma cases

Table 5: Genes (48) up-regulated in a nontumor tissue in an early recurrence group of type B hepatocellular carcinoma cases

Table 6: Genes (12) up-regulated in a nontumor tissue in an early recurrence group of type C hepatocellular carcinoma cases

Table 7: Genes (75) up-regulated in a tumor tissue in an early recurrence group of type B hepatocellular carcinoma cases

Table 8: Genes (38) up-regulated in a tumor tissue in an early recurrence group of type C hepatocellular carcinoma cases

Table 1 Genes (24) up-regulated in nontumor tissue in late recurrence group of hepatitis B cases (BNgood)

No.	Gene	Overlapped group	
1	TNFSF14		
2	MMP2		
3	SAA2	Late recurrence group (type B, tumor)	
4	COL1A1		
5	COL1A2		
6	DPYSL3		
7	PPARD		
8	LUM		
9	MSTP032		
10	CRP		
11	TRIM38		
12	S100A6		
13	PZP		
14	EMP1		
15	AI590053		
16	MAP3K5		
17	TIMP1		
18	GSTM1	Late recurrence group (type B, tumor)	Late recurrence group (type C, tumor)
19	CSDA		
20	GSTM2	Late recurrence group (type B, tumor)	Late recurrence group (type C, tumor)
21	SGK	Late recurrence group (type B, tumor)	
22	LMNA		
23	MGP		
24	LTBP2		

Table 2 Genes (10) up-regulated in nontumor tissue in late recurrence group of hepatitis C cases (CNgood)

No.	Gene	Overlapped group	
25	M10098	Late recurrence group (type B, tumor)	Late recurrence group (type C, tumor)
26	PSMB8		
27	RALGDS		
28	APOL3		
29	GBP1		
30	RPS14		
31	CXCL9		
32	DKFZp564F212		
33	CYP1B1		
34	TNFSF10		

Table 3 Genes (137) up-regulated in tumor tissue in late recurrence group of hepatitis B cases (BTgood)

No.	Gene	Overlapped group	
35	HP		
25	M10098	Late recurrence group (type C, tumor)	Late recurrence group (type C, nontumor)
36	CYP2E1		
37	HDL	Late recurrence group (type C, tumor)	
38	GPX4		
39	G0S2		
40	HAO2		
41	ATF5	Late recurrence group (type C, tumor)	
42	MT1F	Late recurrence group (type C, tumor)	
43	CYP3A4	Late recurrence group (type C, tumor)	
44	Scd		
45	SERPINA7		
46	AKR1D1		
47	AL031602		
48	TSC501		
18	GSTM1	Late recurrence group (type B, nontumor)	Late recurrence group (type C, tumor)
3	SAA2	Late recurrence group (type B, nontumor)	
49	BHMT	Late recurrence group (type C, tumor)	
50	HADHSC		
51	FBXO9		
52	KIAA0442		
53	KIAA0293	Late recurrence group (type C, tumor)	
54	IGHG3		
55	ADH2	Late recurrence group (type C, tumor)	
20	GSTM2	Late recurrence group (type B, nontumor)	Late recurrence group (type C, tumor)
56	PPIF		
57	ALDH8A1		
58	IGLJ3		
59	HCN3		
60	ADH6	Late recurrence group (type C, tumor)	
61	AK02720	Late recurrence group (type C, tumor)	
62	NET-6		
63	CYP2D6		
64	MAFB		
65	GHR		
66	KHK		
67	ADFP		
68	LCE		
69	MPDZ	Late recurrence group (type C, tumor)	
70	TEM6		
71	KIAA0914		
72	KLKB1		
73	M11167	Late recurrence group (type C, tumor)	
21	SGK	Late recurrence group (type B, nontumor)	
74	EHHADH		
75	MBL2	Late recurrence group (type C, tumor)	
76	APP		
77	MT1G		
78	TPD52L1	Late recurrence group (type C, tumor)	
79	CXCL10		
80	AI972416		
81	FCGR2B		
82	IGL@		
83	FLJ10134		
84	PPAP2B		
85	CDC42		
86	HBA2		
87	CYP1A2	Late recurrence group (type C, tumor)	
88	CYP2B6		
89	DKFZP586B1621		
90	MTP		
91	X07868		
92	RNAHP	Late recurrence group (type C, tumor)	
93	HLF	Late recurrence group (type C, tumor)	
94	PPP1R3C		
95	CDC2L2		
96	NRIP1		
97	GPD1		

(Table 3, continued)

No.	Gene	Overlapped group	
98	KIAA1053		
99	CCL19		
100	CRI1		
101	THBS1		Late recurrence group (type C, tumor)
102	SLC5A3		
103	GADD45B		
104	AGL		
105	ADK		
106	IGKC		
107	CYP2A6		Late recurrence group (type C, tumor)
108	GADD45A		Late recurrence group (type C, tumor)
109	FLJ20701		
110	LOC57826		
111	SLC2A2		
112	CIRBP		
113	CGI-26		
114	DEFB1		
115	HMGCS1		
116	ODC1		
117	GLUL	Early recurrence group (type B, nontumor)	Late recurrence group (type C, tumor)
118	CYP27A1		
119	SULT2A1		Late recurrence group (type C, tumor)
120	AK024828		
121	PHLDA1		
122	NR1I2		
123	MSRA		
124	RNASE4		
125	AI339732		
126	HBA2		
127	AL050025		
128	CSAD		
129	SID6-306		
130	NM024561		
131	BCKDK		
132	SLC6A1		
133	CG018		
134	GNE		
135	CKLFSF6		
136	COMT		
137	AL135960		
138	KIAA0179		
139	c-maf		
140	OSBPL11		
141	R06655		Late recurrence group (type C, tumor)
142	KIAA04461		
143	IGF1		Late recurrence group (type C, tumor)
144	HBA1		
145	LOC55908		
146	ENPEP		
147	TXNIP		
148	KIAA0624		
149	ENPP1		
150	CYP4F3		
151	CAV2		
152	BE908931		
153	LECT2		
154	MLLT2		
155	FLR1		
156	TF		
157	DAO		
158	AI620911		
159	GBP1		
160	UGP2		
161	GADD45B		
162	SC4MOL		
163	BE908931		
164	TUBB		
165	EPHX2		
166	SORD		

Table 4 Genes (104) up-regulated in tumor tissue in late recurrence group of hepatitis C cases (CTgood)

No.	Gene	Overlapped group		
167	LEAP-1			
168	PPD			
37	HDL		Late recurrence group (type B, tumor)	
43	CYP3A4		Late recurrence group (type B, tumor)	
107	CYP2A6		Late recurrence group (type B, tumor)	
25	M10098	Late recurrence group (type C, nontumor)	Late recurrence group (type B, tumor)	
169	RACE			
170	SLC27A5			
171	FLJ20581			
172	FLJ10851			
53	KIAA0293		Late recurrence group (type B, tumor)	
173	C9			
174	AL354872			
175	AKR1C1			
176	PCK1			
18	GSTM1		Late recurrence group (type B, tumor)	Late recurrence group (type B, nontumor)
87	CYP1A2		Late recurrence group (type B, tumor)	
177	ANGPTL4			
178	AOX1			
179	SDS			
20	GSTM2		Late recurrence group (type B, tumor)	Late recurrence group (type B, nontumor)
73	M11167		Late recurrence group (type B, tumor)	
180	CYP2C9			
181	SIPL			
182	GLYAT			
75	MBL2		Late recurrence group (type B, tumor)	
183	CYP1A1			
184	CRP			
141	R06655		Late recurrence group (type B, tumor)	
185	ACADL			
93	HLF		Late recurrence group (type B, tumor)	
186	NR1I3			
187	CA2			
188	CYP2C8			
189	PON1			
55	ADH2		Late recurrence group (type B, tumor)	
92	RNAHP		Late recurrence group (type B, tumor)	
190	AQP9			
119	SULT2A1		Late recurrence group (type B, tumor)	
191	SPP1			
192	KIAA0934			
193	AKAP12			
194	APOF			
195	FMO3			
196	SLC22A1			
197	DCXR			
198	CYP3A7			
199	SOCS2			

(Table 4, continued)

No.	Gene		Overlapped group
101	THBS1		Late recurrence group (type B, tumor)
41	ATF5		Late recurrence group (type B, tumor)
200	BCRP		
60	ADH6		Late recurrence group (type B, tumor)
201	humNRDR		
202	GADD45G		
203	SRD5A1		
204	ABCA8		
61	AK026720		Late recurrence group (type B, tumor)
205	APOC4		
206	FTHFD		
207	ISG15		
208	IGFBP2		
49	BHMT		Late recurrence group (type B, tumor)
209	DNASE1L3		
210	SRD5A1		
211	E2IG4		
212	COL1A2		
213	C20orf46		
214	ESR1		
215	BLVRB		
216	LRP16		
217	SLC1A1		
218	ABCB6		
69	MPDZ		Late recurrence group (type B, tumor)
219	FBP1		
220	ALAS1		
221	IFIT1		
222	PPARGC1		
223	Id-1H		
224	RBP1		
225	CSHMT		
226	LOC155066		
42	MT1F		Late recurrence group (type B, tumor)
227	AGXT2L1		
228	TIMM17A		
229	SEC14L2		
230	MAOA		
231	MYC		
232	ACAA2		
233	AL109671		
234	ABCA6		
143	IGF1		Late recurrence group (type B, tumor)
235	GRHPR		
236	HADH2		
237	AFM		
238	COL1A1		
239	MTHFD1		
240	NMT2		
108	GADD45A		Late recurrence group (type B, tumor)
241	UGT2B15		
242	AR		
78	TPD52L1		Late recurrence group (type B, tumor)
243	sMAP		
117	GLUL	Early recurrence group (type B, nontumor)	Late recurrence group (type B, tumor)
244	dJ657E11.4		

Table 5 Genes (48) up-regulated in nontumor tissue in early recurrence group of hepatitis B cases (BNbad)

No.	Gene	Overlapped group	
245	CTH		Early recurrence group (type B, tumor)
246	OAT		
247	PRODH		Early recurrence group (type B, tumor)
248	CYP3A7		
249	DDT		Early recurrence group (type B, tumor)
250	PGRMC1		
251	AKR1C1		
252	HGD		Early recurrence group (type B, tumor)
253	FHR-4		
254	AL354872		
255	FST		Early recurrence group (type B, tumor)
256	COX4		
257	APP		
258	PSPHL		
259	CYP1A1		
260	ZNF216		
261	LEPR		Early recurrence group (type B, tumor)
262	TOM1L1		
263	PECR		
264	ALDH7A1		
265	GNMT		
266	OATP-C		
267	AKR1B10	Early recurrence group (type C, nontumor)	Early recurrence group (type B, tumor)
268	ANGPTL3		
269	AASS		
270	CALR		
271	BAAT		
272	PMM1		
273	RAB-R		
117	GLUL	Late recurrence group (type C, tumor)	Late recurrence group (type B, tumor)
274	CSHMT		
275	UGT1A3		
276	HSPG1		
277	QPRT	Early recurrence group (type C, nontumor)	
278	DEPP		
279	CA2		Early recurrence group (type B, tumor)
280	FTHFD		
281	LAMP1		
282	FKBP1A		
283	BNIP3		
284	MAP3K12		
285	ASS		Early recurrence group (type B, tumor)
286	ACTB		
287	PLAB		Early recurrence group (type B, tumor)
288	ENO1L1		
289	IGFBP3		
290	UK114		
291	ERF-1		

Table 6 Genes (12) up-regulated in nontumor tissue in early recurrence group of hepatitis C cases (CNbad)

No.	Gene	Overlapped group	
292	ALB		
293	NR0B2		
267	AKR1B10	Early recurrence group (type B, nontumor)	Early recurrence group (type B, tumor)
294	MAFB		
295	BF530535		
296	MRPL24		
297	DSIPI		
277	QPRT	Early recurrence group (type B, nontumor)	
298	VNN1		
299	IRS2		
300	FMO5		
301	DCN		

Table 7 Genes (75) up-regulated in tumor tissue in early recurrence group of hepatitis B cases (BTbad)

No.	Gene	Overlapped group	
247	PRODH	Early recurrence group (type B, nontumor)	
302	PLA2G2A		Early recurrence group (type C, tumor)
303	SDS		
304	LGALS3BP		
305	BACE2		
261	LEPR	Early recurrence group (type B, nontumor)	
306	RCN1		
307	MRC1		
308	TM4SF5		
309	NK4		
310	PABL		
311	IGFBP2		
312	GRINA		
313	IFI27		
314	GP2		
315	GA		
316	P4HA2		
317	KYNU		
318	PCK1		
319	UQBP		
320	HLA-DRB1		
252	HGD	Early recurrence group (type B, nontumor)	
321	HTATIP2		
322	GGT1		
323	CTSH		
324	MVP		
325	SLC22A1L		
326	GMNN		
327	COM1		
328	TM7SF2		
245	CTH	Early recurrence group (type B, nontumor)	
329	KDELR3		
330	VPS28		
279	CA2	Early recurrence group (type B, nontumor)	
331	SFN		
332	NM023948		
333	OPLAH		

(Table 7, continued)

No.	Gene	Overlapped group		
334	DGCR6			
335	INSIG1			
267	AKR1B10	Early recurrence group (type B, nontumor)		Early recurrence group (type C, nontumor)
336	PTGDS		Early recurrence group (type C, tumor)	
337	SLC25A15			
338	SEPW1			
339	CD9			
340	UQCRB			
285	ASS	Early recurrence group (type B, nontumor)		
341	CPT1A			
287	PLAB	Early recurrence group (type B, nontumor)		
342	GPAA1			
343	HF1			
344	GPX2			
345	COPEB			
346	NDRG1			
347	SYNGR2			
348	GOT1			
349	POLR2K			
350	AATF			
255	FST	Early recurrence group (type B, nontumor)		
351	OAZIN			
352	RPL7			
353	KIAA0128			
354	CLDN7			
355	ABCB6			
356	GK			
357	LU		Early recurrence group (type C, tumor)	
358	TNFSF4			
359	OSBPL9			
360	GSN			
361	LGALS4			
249	DDT	Early recurrence group (type B, nontumor)		
362	EIF3S3			
363	SLC12A2			
364	RAMP1			
365	HSPB1			
366	AI201594			

Table 8 Genes (38) up-regulated in tumor tissue in early recurrence group of hepatitis C cases (CTbad)

No.	Gene	Overlapped group
367	BL34	
368	AL022324	
369	IGHM	
370	TXNIP	
371	FSTL3	
372	AW978896	
373	NM018687	
374	L48784	
375	AJ275355	
376	PER1	
377	CYBA	
302	PLA2G2A	Early recurrence group (type B, tumor)
378	SGK	
379	FKBP11	
380	AI912086	
381	IGLJ3	
382	IGKC	
336	PTGDS	Early recurrence group (type B, tumor)
383	M20812	
384	AGRN	
385	IL2RG	
386	X07868	
387	PKM2	
388	FGFR3	
389	TRB@	
390	TNFAIP3	
391	TTC3	
392	LPA	
393	AL049987	
394	IER5	
395	BSG	
396	TM4SF3	
397	HMGB2	
357	LU	Early recurrence group (type B, tumor)
398	CCL19	
399	PAM	
400	PIK3R1	
401	RANGAP1	

In Table 5, "CTH" and "AL354872" are genes, which encode the same protein.

The above-described genes can be included in a kit for evaluating cancer, singly or in combination, as appropriate. Examples of a gene set consisting of several genes may include those shown in Table 16 (described later). The above genes may have the partial sequence thereof. Such genes can be used as probes for detecting the expression

of the genes shown in the table.

Moreover, the kit of the present invention may comprise primers used for gene amplification, a buffer solution, polymerase, etc.

With regard to such primers used for gene amplification, the DNA sequence and mRNA sequence of each gene sequence are obtained from database, and in particular, information including the presence or absence of a variant and exon-intron structure is obtained. The same sequences as sequences of portions corresponding to coding regions are used as target. One primer is intended to bridge over an adjacent exon, and it is designed such that only mRNA is detected. Otherwise, primer candidates are obtained using the web software "Primer3" (provided by Steve Rozen and Whitehead Institute for Biomedical Research), and thereafter, homology search is carried out using BLAST (NCBI) search, so as to select primers, which are able to avoid miss-annealing to similar sequences.

The sequence numbers of preferred primers are represented by the general formulas 2n-1 and 2n (wherein n represents an integer between 1 and 114). In the present invention, a primer represented by 2n-1 and a primer represented by 2n can be used as a set of primers. For example, when n is 1, a primer set consisting of the primers shown in SEQ ID NOS: 1 and 2 can be used, and when n is 2, a primer set consisting of the primers shown in SEQ ID NOS: 3 and 4 can be used. Particularly preferred primers can be obtained, when n is 2, 4, 7, 9, or 17.

Moreover, in (1) above, it is also possible to carry out the quantitative analysis of gene expression via immuno-dot blot assay or immunostaining. Such immuno-dot blot assay or immunostaining can be carried out according to common methods using an antibody reacting with the expression products of the genes shown in Tables 1 to 8. As such an antibody, a commercially available antibody may be used, or an antibody obtained by immunization of animals such as a mouse, a rat, or a rabbit, may also be used.

The present invention will be more specifically described in the following examples. However, these examples are not intended to limit the technical scope of the present invention.

Example 1

Detection of up-regulated gene in hepatocellular carcinoma cases

As described below, using human hepatic tissues obtained from type B and type C hepatocellular carcinoma cases, molecules for suppressing the recurrence of hepatocellular carcinoma were identified at a gene level.

In order to understand a recurrence mechanism occurring after an operation to resect hepatocellular carcinoma and determine a gene capable of predicting the presence or absence of recurrence, gene expression profile analysis was carried out, using several cases, the recurrence periods of which were different. 51 cases, which were at stages I and II based on TNM classification, were used as targets. 5 cases wherein the cancer had not recurred for 4 or more years after the operation, and 5 cases wherein the cancer had recurred within 1 year after the operation, were selected. Thereafter, expression analysis was carried out using an HG-U133A array manufactured by Affymetrix.

The TRIzol reagent (Life Technologies, Gaithersburg, MD) was added to frozen tissues, and the obtained mixture was then homogenated with Polytron. Thereafter, chloroform was added to the homogenate, and they were then fully mixed, followed by centrifugation. After completion of the centrifugation, the supernatant was recovered, and an equivalent amount of isopropanol was added thereto. Thereafter, the precipitate of total RNA was recovered by centrifugation.

Type B hepatocellular carcinoma cases (wherein the causal virus is a hepatitis B virus) were divided into the following groups: the nontumor tissues and tumor tissues of 2 early recurrence cases; and the nontumor tissues and tumor tissues of 2 late recurrence cases. Also, type C hepatocellular carcinoma cases (wherein the causal virus is a hepatitis C virus) were divided into the following groups: the nontumor tissues and

tumor tissues of 3 early recurrence cases; and the nontumor tissues and tumor tissues of 3 late recurrence cases. Thus, the total 8 groups were subjected to expression analysis.

For each sample group, 15 μg of total RNA was prepared. Thereafter, biotin-labeled cRNA was synthesized based on GeneChip Expression Analysis Technical Manual by Affymetrix. Using T7-$(dt)_{24}$ primer and Superscript II reverse transcriptase (Invitrogen Life Technology), the reaction was carried out for 1 hour, so as to synthesize first strand cDNA. Thereafter, *E. coli* DNA ligase, *E. coli* DNA polymerase, and *E. coli* RNase H were added thereto, and the obtained mixture was then allowed to react at 16°C for 2 hours. Finally, T4 DNA polymerase was added to the reaction product, so as to synthesize double strand cDNA. After cleanup of the cDNA, the BioArray high yield RNA transcript labeling kit (Affymetrix, Inc, CA) was used for *in vitro* transcription at 37°C for 4 hours, so as to synthesize biotin-labeled cRNA. A hybridization probe solution was prepared based on the Technical Manual, and the above solution was then added to GeneChip HG-U133A (Affymetrix, Inc, CA; containing 22,283 human genes), obtained by pre-hybridization at 45°C for 45 minutes. Thereafter, hybridization was carried out at 45°C for 16 hours. Thereafter, the reaction product was washed with GeneChip Fluidics Station 400 (Affymetrix, Inc, CA), and was then stained with streptavidin phycoerythrin and biotinylated antistreptavidin. Thereafter, the resultant was subjected to scanning using an HP GeneArray scanner (Affymetrix, Inc, CA).

The obtained data was analyzed using GeneSpring ver.5.0 (SiliconGenetics, Redwood, CA). After completion of normalization, using the signal of the control gene BioB used for intrinsic quantification as a detection limit (corresponding to several copies per cell). A gene, which has a signal intensity of 100 or greater and also has a present flag in at least one chip, was defined as a target of the analysis. As a result, 7,444 genes were determined to be such analysis targets. In nontumor tissues, genes having 2.5 times or more difference between the early recurrence group and the late recurrence group have been identified. In tumor tissues, genes having 3 times or more difference between such two groups have been identified.

As a result, among the selected 7,444 genes, genes having 2.5 times or more difference between the absence and the presence of recurrence in nontumor tissues consisted of 34 up-regulated genes and 58 down-regulated genes. On the other hand, genes having 3 time or more difference between such two groups in tumor tissues consisted of 215 up-regulated genes and 110 down-regulated genes. Among these genes, as a gene up-regulated in the recurrence-absent group in both cases of type B and type C, no such genes were found in nontumor tissues, whereas 26 genes were found in tumor tissues. On the other hand, among these genes, as a gene up-regulated in the recurrence-present group in both cases of type B and type C, 2 genes were found in nontumor tissues, whereas 3 genes were found in tumor tissues. Moreover, there were genes up-regulated in both tumor and nontumor tissue. There were found 5 genes up-regulated in the recurrence-absent group, and 10 genes up-regulated in the recurrence-present group (Table 9).

It is to be noted that the total is not 402 but 401 in Table 9. This is because the overlapping of GLUL is a particular case.

Table 9 Genes associated with recurrence of hepatocellular carcinoma

	Up-regulated in late recurrence group		Up-regulated in early recurrence group		Both cases
	nontumor tissue	tumor tissue	nontumor tissue	tumor tissue	
Hepatitis B	24	137	48	75	4 / 10
Hepatitis C	10	104	12	38	1 / 0
Both types	0	26	2	3	
Total	34	215	58	110	244 / 158

Total 401

From the results shown in Table 9, it can be said that with regard to a difference in recurrence prognosis, a change in gene expression is greater in a tumor tissue than in a nontumor tissue, and that such a change in gene expression is greater in type B hepatocellular carcinoma cases than in type C hepatocellular carcinoma cases. In addition, there are genes associated with recurrence prognosis, which are found independently of a causal virus, but unexpectedly, such genes are rare. As in the case of the development of cancer, it is considered that different mechanisms are involved in the recurrence of cancer, depending on the type of a causal virus.

In the analysis of a sample phylogenetic tree, the expression profiles of all genes are first divided into nontumor tissues and tumor tissues. In each of such nontumor tissues and tumor tissues, a genetic affiliation, which is not caused by recurrence prognosis but caused by a causal virus, was observed (Figure 1). In Figure 1, with regard to notation indicating each test group, such as "BNbad" or "BNgood," the first alphabet indicates the type of a virus. That is, "B" represents hepatitis B virus, and "C"

represents hepatitis C virus. The second alphabet "N" represents a nontumor tissue, and "T" represents a tumor tissue. Moreover, "bad" represents early recurrence, and "good" represents late recurrence.

It is considered that gene expression affecting recurrence prognosis is caused by a change in the gene expression of limited genes.

As stated above, candidate genes capable of clarifying a recurrence mechanism or predicting the presence or absence of recurrence were found (Tables 1 to 8).

Example 2

Study of correlation between the recurrence period and an expression level of genes in each group in type C hepatocellular carcinoma cases

As mentioned below, with regard to genes up-regulated in the nontumor tissues of a late recurrence group and an early recurrence group in type C hepatocellular carcinoma cases, the correlation between the recurrence period and an expression level was studied.

The total 22 nontumor tissue samples, including 6 cases of type C hepatocellular carcinoma used in the gene expression profile analysis, were used as targets. The clinicopathological findings of each case and the recurrence period (that is, the period of time in which the cancer has not yet recurred) are shown in Table 10A.

Table 10A Type C hepatocellular carcinoma cases

Case No.	Sex	Age	Nontumor tissue	stage	Number of months without recurrence	Microarray
59	M	66	CH	I	84	Late recurrence group
18	M	68	LC	I	58	Late recurrence group
6	M	65	CH	II	51	Late recurrence group
25	M	51	CH	I	45	
29	M	70	CH	II	43	
12	M	66	CH	II	41	
4	M	65	CH	I	40	
48	F	65	LC	I	39	
31	M	60	LC	I or II	38	
16	M	70	CH	I	37	
22	M	65	CH	I	34	
3	F	71	LC	I	29	
65	M	60	LC	I	29	
30	F	62	LC	II	28	
10	M	56	LC	I	26	
23	M	62	CH	II	16	
26	M	70	LC	I	16	
14	M	62	CH	II	14	Early recurrence group
62	M	66	LC	I	13	
17	M	54	LC	I	12	
15	F	68	LC	II	8	Early recurrence group
44	M	58	CH	I	4	Early recurrence group

CH: chronic hepatitis; LC: liver cirrhosis
Stage of case 31: undetermined
The term "number of months without recurrence" includes not only the number of months required for recurrence, but also includes the investigation period in which recurrence was not observed.

In addition, the cases shown in Table 10A were changed or revised as a result of follow-up study. Moreover, with regard to the total 35 cases, including cases added as the targets of the present example, the clinicopathological findings of each case and the recurrence period (that is, the period of time in which the cancer has not yet recurred) are shown in Table 10B.

Table 10B Type C hepatocellular carcinoma cases

Case No.	Sex	Age	Nontumor tissue	stage	Number of months without recurrence	Microarray
59	M	66	CH	I	>94	Late recurrence group
6	M	65	CH	II	65	Late recurrence group
25	M	51	CH	I	> 58	
18	M	68	LC	I	58	Late recurrence group
12	M	66	CH	II	41	
4	M	65	CH	I	>40	
29	M	70	CH	II	39	
16	M	70	CH	I	>37	
48	F	65	LC	I	37	
31	M	60	LC	I	37	
80	M	73	CH	II	34	
22	M	65	CH	I	33	
3	F	71	LC	I	29	
65	M	60	LC	I	28	
30	F	62	LC	II	26	
10	M	56	LC	I	25	
70	M	57	LC	II	24	
79	M	73	LC	I	22	
73	M	50	CH	II	20	
81	F	69	LC	I	17	
26	M	70	LC	I	16	
72	M	71	LC	II	16	
69	M	66	LC	II	15	
14	M	62	CH	II	14	Early recurrence group
78	F	66	CH	I	13	
82	M	71	CH	I	13	
17	M	54	LC	I	12	
71	M	57	LC	II	12	
77	F	65	LC	I	10	
62	M	66	LC	I	9	
74	M	67	CH	II	9	
15	F	68	LC	II	8	Early recurrence group
76	M	72	NL	I	7	
75	M	65	CH	II	6	
44	M	58	CH	I	4	Early recurrence group

CH: chronic hepatitis; LC: liver cirrhosis; NL: normal liver

The term "number of months without recurrence" includes not only the number of months required for recurrence, but also includes the period in which recurrence has not yet been observed at the time of investigation.

With regard to the total 21 genes consisting of 9 genes (CNgood) up-regulated in the nontumor tissues of the late recurrence group shown in Table 2 and 12 genes (CNbad) up-regulated in the nontumor tissues of the early recurrence group shown in Table 6, the relationship between the recurrence period and an expression level was

analyzed.

First, total RNA was extracted from the nontumor liver tissue of each case by the same method as that described in Example 1 above.

In order to eliminate the influence of DNA mixed therein, the total RNA was treated with DNase I (DNase I, TAKARA SHUZO, Kyoto, Japan) at 37°C for 20 minutes, and it was then purified again with a TRIzol reagent. Using 10 μg of the total RNA, a reverse transcription reaction was carried out with 100 μl of a reaction solution comprising 25 units of AMV reverse transcriptase XL (TAKARA) and 250 pmol of a 9-mer random primer.

Real-time PCR was carried out using 0.25 to 50 ng each of synthetic cDNA. 25 μl of a reaction solution, SYBR Green PCR Master mix (Applied Biosystems, Foster City, CA) was used, and ABI PRISM 7000 (Applied Biosystems) was employed. PCR was carried out under conditions wherein preliminary heating was carried out at 95°C for 10 minutes, and thereafter, a cycle consisting of 95°C for 15 seconds and 60°C (or 65°C) for 60 seconds, was repeated 40 to 45 times.

Using glyceraldehyde 3-phosphatase dehydrogenase (GAPDH) or 18S rRNA as an internal standard gene of each sample, relative quantitative analysis, and partially, absolute quantitative analysis, were carried out. Values obtained by subjecting standard samples to serial dilution and simultaneous measurement, were used to produce a calibration curve. A threshold line for optimization of such a calibration curve was determined, and the number of threshold PCR cycles, a threshold cycle value (Ct) was then obtained for each sample. A Δ Ct value was obtained by subtracting the Ct value of GAPDH or 18S rRNA from the Ct value of a target gene, and the obtained value was defined as the relative expression level of the target gene. Moreover, values obtained using the formula ($2^{(-\Delta Ct)}$) were used for evaluation of a linear expression level.

On the other hand, with regard to genes whose absolute expression level can be calculated relative to a calibration curve, the absolute expression level of a target gene and that of an internal standard gene were obtained. Thereafter, the ratio of the target

gene expression level/the internal standard gene expression level was calculated for each sample, and it was used for evaluation. All such measurements were carried out in a duplicate manner.

In Tables 11A, 11B, 12A, and 12B, the term "correspondence with microarray" is used to mean that when the ratio between the late recurrence group (case Nos. 59, 18, and 6) and the early recurrence group (case Nos. 14, 15, and 44) was obtained from the results of quantitative PCR performed on 6 cases (case Nos. 59, 18, 6, 14, 15, and 44 in Table 10A or 10B) used in the microarray analysis, genes, the above ratio of which was 1.5 or greater, corresponded with the results of the microarray in Example 1. Genes corresponding with the microarray results were indicated with the mark O. The above ratio is 1.5 or greater, and preferably 2 or greater. The number in the parenthesis adjacent to the mark O indicates such a ratio (the average ratio of 3 cases). The mark X in the "correspondence with microarray" column indicates a gene that does not correspond with the microarray results. The mark XX indicates a gene, which exhibits an opposite correlation with the microarray results.

In Tables 11A, 11B, 12A, and 12B, the term "correlation" is used to mean a correlation between the gene expression level and the recurrence period in 22 cases, or in 31 cases wherein the number of months in which the recurrence of the cancer had occurred was determined. In the case of a significant correlation, O or the r value was indicated, and further, the p value was also indicated.

In Tables 11B and 12B, with regard to genes exhibiting a significant difference in expression levels between 19 cases of the recurrence within 24 months, and 6 cases of no recurrence for 40 months or more (the upper case of the "significant difference between two groups" column in Tables 11B and 12B) or 4 cases of no recurrence for 58 months or more (the lower case of the "significant difference between two groups" column in Tables 11B and 12B), p values (Mann-Whitney U test) were shown in the "significant difference between two groups" column.

Primer sequences (sense strand (forward), antisense strand (reverse)) used for

the test are shown in Tables 11A, 11B, 12A, and 12B (SEQ ID NOS: 1 to 88).

The results obtained by analyzing the 9 gene candidates (CNgood) up-regulated in nontumor tissues in the late recurrence group of type C hepatocellular carcinoma cases are shown in Tables 11A and 11B. Table 11A shows the analysis results obtained by quantitative PCR, which was performed on the cases shown in Table 10A as targets, under the conditions shown in Table 11A using GAPDH as an internal standard gene.

Table 11A Results of quantitative PCR of "genes up-regulated in nontumor tissues in late recurrence group of hepatitis C cases"

No.	Gene	Forward/ reverse	Primer sequence (5'-3')	SEQ ID NO.	Annealing temperature	Correspondence with microarray	Correlation
26	PSMB8	F	AGACTGTCAGTACTGGGAGC	1	60°C	O(2.52)	
		R	GTCCAGGACCCTTCTTATCC	2			
27	RALGDS	F	GACGTGGGAAGACGTTTCCA	3	60°C	O(4.13)	O(p=0.0118)
		R	TGGATGATGCCCGTCTCCTT	4			
28	APOL3	F	AATTGCCCAGGGATGAGGCA	5	60°C	O(2.69)	
		R	TGGACTCCTGGATCTTCCTC	6			
29	GBP1	F	GAGAACTCAGCTGCAGTGCA	7	65°C	O(6.00)	O(p=0.0031)
		R	TTCTAGCTGGGCCGCTAACT	8			
30	RPS14	F	GACGTGCAGAAATGGCACCT	9	60°C	× (0.96)	
		R	CAGTCACACGGCAGATGGTT	10			
31	CXCL9	F	CCTGCATCAGCACCAACCAA	11	65°C	O(11.5)	
		R	TGGCTGACCTGTTTCTCCCA	12			
32	DKFZp564F212	F	CCACATCCACCACTAGACAC	13	60°C	O(4.75)	O(p=0.0541)
		R	TGACAGATGTCCTCTGAGGC	14			
33	CYP1B1	F	CCTCTTCACCAGGTATCCTG	15	60°C	O(2.33)	
		R	CCACAGTGTCCTTGGGAATG	16			
34	TNFSF10	F	GCTGAAGCAGATGCAGGACA	17	60°C	O(2.50)	O(p=0.0424)
		R	CTAACGAGCTGACGGAGTTG	18			

With regard to "correspondence with microarray," the ratio of late recurrence group and early recurrence group was obtained from the results of quantitative PCR performed on 6 cases used in microarray analysis, and genes with the ratio of 1.5 or greater were indicated with O.

With regard to "correlation," genes exhibiting correlation between the gene expression levels of 22 cases and the period of time required for recurrence were indicated with O, and the p values thereof were also shown.

As a result, it was found that 8 genes corresponded with the microarray results, and that among such genes, 4 genes (RALGDS, GBP1, DKFZp564F212, and TNFSF10) exhibited a correlation with the recurrence period.

Likewise, Table 11B shows the analysis results obtained by quantitative PCR, which was performed on the 10 genes shown in Table 11B and the cases shown in Table 10B as targets, under the conditions shown in the table using GAPDH or 18S rRNA as an internal standard gene.

Table 11B Results of quantitative PCR of "genes up-regulated in nontumor tissues in late recurrence group of hepatitis C cases"

No.	Gene	Forward/ reverse	Primer sequence (5'-3')	SEQ ID NO.	Annealing temperature	Correspondence with microarray, normalized with GAPDH	Correspondence with microarray, normalized with 18S rRNA	Correlation (GAPDH)	Correlation (18S rRNA)	Significant difference between two groups (GAPDH)	Significant difference between two groups (18S rRNA)
1	M10098	F	GGAGGTTCGAAGACGATCAG	19	65°C	× ×(0.60)	-				
		R	GTGGTGCCCTTCCGTCAATT	20							
2	PSMB8	F	AGACTGTCAGTACTGGGAGC	21	60°C	O(1.92)	O(3.60)		r=0.421 (p=0.0177)		
		R	GTCCAGGACCCTTCTTATCC	22							
3	RALGDS	F	GTGTGGCCAACTGTGTCATC	23	65°C	O(6.71)	O(8.23)		r=0.377 (p=0.0361)	0.0314	
		R	CTTCAGACGGTGGATGGAGT	24							
4	APOL3	F	AATTGCCCAGGGATGAGGCA	25	60°C	O(1.65)	O(2.13)				
		R	TGGACTCCTGGATCTTCCTC	26							
5	GBP1	F	AACAAGCTGGCTGGAAAGAA	27	65°C	O(6.87)	O(5.76)	r=0.359 (p=0.0469)	r=0.374 (p=0.0377)		
		R	GTACACGAAGGTGCTGCTCA	28							
6	RPS14	F	GACGTGCAGAAATGGCACCT	29	60°C	O(2.02)	O(3.35)	r=0.383 (p=0.0329)	r=0.458 (p=0.0089)		0.0357
		R	CAGTCACACGGCAGATGGTT	30							
7	CXCL9	F	CCTGCATCAGCACCAACCAA	31	65°C	O(14.3)	O(12.5)	r=0.392 (p=0.0282)	r=0.437 (p=0.0132)		0.0131
		R	TGGCTGACCTGTTTCTCCCA	32							
8	DKFZp564F212	F	TGGGCAAGTGAGGTCTTCTT	33	60°C	O(4.69)	O(8.40)		r=0.501 (p=0.0036)	0.0485 0.0094	0.0075 0.0074
		R	CTGAGGATCACTGGTATCGC	34							
9	CYP1B1	F	GACCCCCAGTCTCAATCTCA	35	65°C	O(4.29)	O(4.78)	r=0.424 (p=0.0167)	r=0.553 (p=0.001)	0.0417 0.0045	0.0042 0.0094
		R	AGTCTCTTGGCGTCGTCAGT	36							
10	TNFSF10	F	GCTGAAGCAGATGCAGGACA	37	60°C	O(3.71)	O(4.54)	r=0.460 (p=0.0085)	r=0.603 (p=0.0002)		0.0062 0.0426
		R	CTAACGAGCTGACGGAGTTG	38							
	GAPDH	F	GGTCGGAGTCAACGGATTTG	39	60°C						
		R	GGATCTCGCTCCTGGAAGAT	40							

The expression level of each gene was evaluated by quantitative PCR using GAPDH as a control gene and was expressed as a relative value to the expression level of the control gene.

With regard to "correspondence with microarray," the ratio of the late recurrence group and the early recurrence group was obtained from the results of quantitative PCR on 6 cases used for microarray analysis, and genes with the ratio of 1.5 or greater were indicated with O.

With regard to "correlation," genes exhibiting a correlation between the gene expression levels of 31 cases wherein the number of months of recurrence had been determined, and the period required for recurrence, were indicated with the r value and the p value.

In "significant difference between two groups," with regard to genes exhibiting a significant difference in expression levels between 19 cases of the recurrence within 24 months, and 6 cases of no recurrence for 40 months or more (the upper case) or 4 cases of no recurrence for 58 months or more (the lower case), p values were indicated (Mann-Whitney U test).

As a result, it was found that when GAPDH was used as an internal standard gene, all the 9 gene candidates exhibiting up-regulation in the late recurrence group corresponded with the microarray results, and that among such genes, 5 genes exhibited a correlation with the recurrence period. In addition, when 18S rRNA was used as an internal standard gene also, all the above 9 gene candidates corresponded with the microarray results, and among them, 8 genes exhibited a correlation with the recurrence period.

A significant difference test was carried out on two groups, the late recurrence group and the early recurrence group. As a result, it was found that when GAPDH was used as a standard gene, 3 genes exhibited a significant difference, and that when 18S rRNA was used as a standard gene, 5 genes exhibited a significant difference.

Subsequently, the results obtained by analyzing the 12 gene candidates (CNbad) up-regulated in nontumor tissues in the early recurrence group of type C hepatocellular carcinoma cases are shown in Tables 12A and 12B. Table 12A shows the analysis

results obtained by quantitative PCR, which was performed on the cases shown in Table 10A as targets, under the conditions shown in Table 12A using GAPDH as an internal standard gene.

Table 12A Results of quantitative PCR of "genes up-regulated in nontumor tissues in early recurrence group of hepatitis C cases"

No.	Gene	F/R	Primer sequence (5'-3')	SEQ ID NO.	Annealing temperature	Correspondence with microarray	Correlation
292	ALB	F	CAAAGCATGGGCAGTAGCTC	41	60°C	O(2.19)	
		R	CAAGCAGATCTCCATGGCAG	42			
293	NR0B2	F	TCTTCAACCCCGATGTGCCA	43	60°C	O(1.48)	
		R	AGGCTGGTCGGAATGGACTT	44			
267	AKR1B10	F	CTTGGAAGTCTCCTCTTGGC	45	60°C	O(2.44)	
		R	ATGAACAGGTCCTCCCGCTT	46			
294	MAFB	F	ACCATCATCACCAAGCGTCG	47	60°C	O(1.56)	
		R	TCACCTCGTCCTTGGTGAAG	48			
295	BF530535	F	GTCGCCTCACCATCTGTACA	49	65°C	O(3.74)	
		R	CTGGAGGACAGCTGCCAATA	50			
296	MRPL24	F	TCCTAGAAGGCAAGGATGCC	51	60°C	×(0.92)	
		R	GTGGGTTTCCTGTCCATAGG	52			
297	DSIPI	F	AACAGGCCATGGATCTGGTG	53	65°C	O(1.85)	
		R	AGGACTGGAACTTCTCCAGC	54			
279	QPRT	F	AGGATAACCATGTGGTGGCC	55	60°C	××(0.413)	O (p=0.0092)
		R	TGCAGCTCCTCTGGCTTGAA	56			
298	VNN1	F	GCTGGAACTTCAACAGGGAC	57	60°C	×(1.11)	
		R	CTGAGGATCACTGGTATCGC	58			
299	IRS2	F	TGAAGCTCAACTGCGAGCAG	59	60°C	O(1.57)	
		R	ACGATTGGCTCTTACTGCGC	60			
300	FMO5	F	ACACAGAGCTCTGAGTCAGC	61	60°C	×(1.13)	
		R	TCCAGGTTAGGAGGGAAGAC	62			
301	DCN	F	CCTCAAGGTCTTCCTCCTTC	63	60°C	×(0.74)	
		R	CACCAGGTACTCTGGTAAGC	64			

QPRT gene is a gene exhibiting an opposite correlation.

As a result, 7 genes corresponded with the microarray results. No genes significantly exhibited a correlation with the recurrence period. However, the QPRT gene significantly exhibited an opposite correlation. Accordingly, this gene was identified as a gene up-regulated in nontumor tissues in the late recurrence group.

Likewise, Table 12B shows the analysis results obtained by quantitative PCR, which was performed on the cases shown in Table 10B as targets, under the conditions shown in Table 12B using GAPDH or 18S rRNA as an internal standard gene.

Table 12B Results of quantitative PCR of "genes up-regulated in nontumor tissues in early recurrence group of hepatitis C cases"

No.	Gene	Forward/ reverse	Primer sequence (5'-3')	SEQ ID NO.	Annealing temperature	Correspondence with microarray, normalized with GAPDH	Correspondence with microarray, normalized with 18S rRNA	Correlation (GAPDH)	Correlation (18S rRNA)	Significant difference between two groups (GAPDH)	Significant difference between two groups (18S rRNA)
1	ALB	F	CAAAGCATGGGCAGTAGCTC	65	60℃	× (1.25)	× × (0.64)				
		R	CAAGCAGATCTCCATGGCAG	66							
2	NR0B2	F	TCTTCAACCCCGATGTGCCA	67	65℃	× (1.13)	× (1.04)				0.0220
		R	AGGCTGGTCGGAATGGACTT	68							
3	AKR1B10	F	CTTGGAAGTCTCCTCTTGGC	69	60℃	× (0.83)	× (0.92)				
		R	ATGAACAGGTCCTCCCGCTT	70							
4	MAFB	F	GACGTGAAGAAGGAGCCACT	71	60℃	× (0.71)	× × (0.61)	r=0.422 (p=0.0171)	r=0.501 (p=0.0036)		0.0281
		R	CGCCATCCAGTACAGATCCT	72							
5	BF530535	F	TGCCATAGTGGCTTGATTTG	73	60℃	× (0.82)	× × (0.48)				0.0486
		R	TCAGAATCCCCATCATCACA	74							
6	MRPL24	F	CAGGGCAAAGTGGTTCAAGT	75	65℃	× × (0.46)	× × (0.31)	r=0.431 (p=0.0147)	r=0.483 (p=0.0053)	0.0083 0.0040	0.0083 0.0426
		R	TCTCAGTGGGTTTCCTGTCC	76							
7	DSIPI	F	AACAGGCCATGGATCTGGTG	77	65℃	O (2.57)	O (1.75)				
		R	AGGACTGGAACTTCTCCAGC	78							
8	QPRT	F	AACTACGCAGCCTTGGTCAG	79	65℃	× (0.72)	× × (0.54)				0.0075
		R	TGGCAGTTGAGTTGGGTAAA	80							0.0231
9	VNN1	F	GCTGGAACTTCAACAGGGAC	81	65℃	× × (0.65)	× × (0.41)			0.0018	0.0009
		R	CTGAGGATCACTGGTATCGC	82						0.0035	0.0074
10	IRS2	F	CCACTCGGACAGCTTCTTCT	83	65℃	× (0.78)	× × (0.63)	r=0.419 (p=0.0181)	r=0.462 (p=0.0082)		
		R	GGATGGTCTCGTGGATGTTC	84							
11	FMO5	F	ACACAGAGCTCTGAGTCAGC	85	60℃	× (1.02)	× × (0.62)				
		R	TCCAGGTTAGGAGGGAAGAC	86							
12	DCN	F	CCTCAAGGTCTTCCTCCTTC	87	60℃	× (1.40)	× (0.77)				
		R	CACCAGGTACTCTGGTAAGC	88							

With regard to "correspondence with microarray," the ratio of the late recurrence group and the early recurrence group was obtained from the results of quantitative PCR on 6 cases used for microarray analysis, and genes with the ratio of 1.5 or greater were indicated with O.

× indicates no difference, and × × indicates an opposite correlation.

With regard to "correlation," genes exhibiting a correlation between the gene expression levels of 31 cases wherein the number of months of recurrence had been determined, and the period required for recurrence, were indicated with the r value (opposite correlation) and the p value.

With regard to "significant difference between two groups," genes exhibiting a significant difference in expression levels between 19 cases of the recurrence within 24 months, and 6 cases of no recurrence for 40 months or more (the upper case) or 4 cases of no recurrence for 58 months or more (the lower case), p values (Mann-Whitney U test) were indicated.

As a result, it was found that when GAPDH or 18S rRNA was used as an internal standard gene, among 12 gene candidates exhibiting up-regulation in the early recurrence group, 1 gene corresponded with the microarray results. However, when GAPDH was used as an internal standard gene, the MAFB gene, the MRPL24 gene, the VNN1 gene, and IRS2 gene significantly exhibited an opposite correlation. In addition, when 18S rRNA was used as an internal standard gene, the NR0B2 gene, the MAFB gene, the BF530535 gene, the MRPL24 gene, the QPRT gene, the VNN1 gene, and the IRS2 gene significantly exhibited an opposite correlation. Accordingly, these genes were identified as genes up-regulated in nontumor tissues in the late recurrence group.

As stated above, as a result of the studies carried out under various conditions, the following 15 genes were identified as genes expressed in nontumor tissues, which can be used for prediction of the recurrence of cancer in type C hepatocellular carcinoma cases: the PSMB8 gene, the RALGDS gene, the GBP1 gene, the RPS14 gene, the

CXCL9 gene, the DKFZp564F212 gene, the CYP1B1 gene, the TNFSF10 gene, the NR0B2 gene, the MAFB gene, the BF530535 gene, the MRPL24 gene, the QPRT gene, the VNN1 gene, and the IRS2 gene. The meanings of the aforementioned genes are as follows:

PSMB8 gene (which is also referred to as LMP7 gene): A proteasome subunit, beta type, 8 gene

RALGDS gene: A ral guanine nucleotide dissociation stimulator gene

GBP1 gene: A guanylate-binding protein 1 gene

RPS14 gene: A ribosomal protein S14 gene

CXCL9 gene: A chemokine (C-X-C motif) ligand 9 gene

DKFZp564F212 gene: An expression gene discovered by German Human Genome Project, whose gene product has not been identified and whose functions have not yet been predicted.

CYP1B1 gene: A cytochrome P450, family 1, subfamily B, polypeptide 1 gene

TNFSF10: An abbreviation of TNF (ligand) super family, member 10, and a TNF-related apoptosis inducing ligand (TRAIL) gene

NR0B2 gene: A nuclear receptor subfamily 0, group B, member 2 gene

MAFB gene: A v-maf musculoaponeurotic fibrosarcoma oncogene homolog B gene

BF530535 gene: A gene whose gene product has not been identified and whose functions have not yet been predicted.

MRPL24 gene: A mitochondrial ribosomal protein L24 gene

QPRT gene: A quinolinate phosphoribosyltransferase gene

VNN1 gene: A vanin 1 gene

IRS2 gene: An insulin receptor substrate 2 gene

Example 3

Study of correlation between the recurrence period and an expression level of genes in each group in type B hepatocellular carcinoma cases

As mentioned below, with regard to genes up-regulated in the nontumor tissues of a late recurrence group and an early recurrence group in type B hepatocellular carcinoma cases, the correlation between the recurrence period and an expression level was studied.

The total 16 nontumor tissue samples, including 4 cases of type B hepatocellular carcinoma used in the gene expression profile analysis, were used as targets. The clinicopathological findings of each case and the recurrence period (that is, the period of time in which the cancer has not yet recurred) are shown in Table 13.

Table 13 Type B hepatocellular carcinoma cases

Case No.	Sex	Age	Nontumor tissue	stage	Number of months without recurrence	Microarray
67	M	45	CH	II	>99	Late recurrence group
87	M	45	CH	I	>92	
85	F	64	NL	II	84	
93	M	58	CH	I	>67	
94	F	59	LC	I	>66	
60	M	60	NL	I	64	Late recurrence group
35	M	69	CH	I	>48	
45	M	68	CH	I	>48	
84	M	51	CH	I/II	47	
54 (86)	M	52	CH	II	27	
47	M	36	CH	I	23	
8	M	68	CH	II	17	
13	F	51	CH	I	14	Early recurrence group
42 (88)	M	74	CH	II	14	
89	M	45	CH	II	9	
9	M	44	CH	II	7	Early recurrence group

CH: chronic hepatitis; LC: liver cirrhosis; NL; normal liver
The term "stage I/II" indicates that it is unknown whether the stage is stage I or II.
The term "number of months without recurrence" includes not only the number of months required for recurrence, but also includes the investigation period in which recurrence was not observed.

With regard to the total 71 genes consisting of 24 genes (BNgood) up-regulated in the nontumor tissues of the late recurrence group shown in Table 1 and 47 genes (BNbad) up-regulated in the nontumor tissues of the early recurrence group shown in Table 5, the relationship between the recurrence period and an expression level was analyzed.

First, total RNA was extracted from the nontumor hepatic tissue of each case by the same method as that described in Example 1 above.

In order to eliminate the influence of DNA mixed therein, the total RNA was treated with DNase I (DNase I, TAKARA SHUZO, Kyoto, Japan) at 37°C for 20 minutes, and it was then purified again with a TRIzol reagent. Using 10 μg of the total RNA, a reverse transcription reaction was carried out with 100 μl of a reaction solution comprising 25 units of AMV reverse transcriptase XL (TAKARA) and 250 pmol of a 9-mer random primer.

Real-time PCR was carried out using 0.25 to 50 ng each of synthetic cDNA. 25 μl of a reaction solution, SYBR Green PCR Master mix (Applied Biosystems, Foster City, CA) was used, and ABI PRISM 7000 (Applied Biosystems) was employed. PCR was carried out under conditions wherein preliminary heating was carried out at 95°C for 10 minutes, and thereafter, a cycle consisting of 95°C for 15 seconds and 60°C (or 65°C) for 60 seconds, was repeated 40 to 45 times.

Using GAPDH or 18S rRNA as an internal standard gene of each sample, absolute quantitative analysis was carried out. Values obtained by subjecting standard samples to serial dilution and simultaneous measurement, were used to produce a calibration curve.

The absolute expression level of a target gene and that of an internal standard gene were obtained. Thereafter, the ratio of the target gene expression level/the internal standard gene expression level was calculated for each sample, and it was used for evaluation. All such measurements were carried out in a duplicate manner.

As with the descriptions in Example 2, the term "correspondence with microarray" shown in Tables 14 and 15 is used to mean that when the ratio of the late recurrence group (case Nos. 67 and 60) and the early recurrence group (case Nos. 13 and 9) was obtained from the results of quantitative PCR performed on 4 cases (case Nos. 67, 60, 13, and 9 in Table 13) used in the microarray analysis, genes, the above ratio of which was 1.5 or greater, corresponded with the results of the microarray in Example 1.

The mark O is given to genes, when the above ratio of is 1.5 or greater, and preferably 2 or greater. The number in the parenthesis adjacent to the mark O indicates the value of such a ratio. The mark X in the "correspondence with microarray" column indicates a gene that does not correspond with the microarray results. The mark XX indicates a gene that exhibits an opposite correlation to the microarray results.

In the "correlation" columns in Tables 14 and 15, with regard to genes, which exhibited a correlation between the gene expression level and the recurrence period in 10 cases wherein the number of months in which the recurrence of the cancer had occurred was determined, the r value and the p value were described.

In the "significant difference between two groups" column in Tables 14 and 15, with regard to genes exhibiting a significant difference in expression levels between 6 cases of the recurrence within 24 months, and 8 cases of no recurrence for 48 months or more (the upper case of the "significant difference between two groups" in Tables 14 and 15) or 6 cases of no recurrence for 60 months or more (the lower case of the "significant difference between two groups" in Tables 14 and 15), p values (Mann-Whitney U test) were indicated.

Primer sequences (sense strand (forward), antisense strand (reverse)) used for the test are shown in Tables 14 and 15 (SEQ ID NOS: 89 to 228).

The results obtained by analyzing the 24 gene candidates (BNgood) up-regulated in nontumor tissues in the late recurrence group of type B hepatocellular carcinoma cases are shown in Tables 14. Table 14 shows the analysis results obtained by quantitative PCR, which was performed on the cases shown in Table 13 as targets, under the conditions shown in Table 14 using GAPDH or 18S rRNA as an internal standard gene.

Table 14 Results of quantitative PCR of "genes up-regulated in nontumor tissues in late recurrence group of hepatitis B cases"

No.	Gene	Forward/ reverse	Primer sequence (5'-3')	SEQ ID NO.	Annealing temperature	Correspondence with microarray, normalized with GAPDH	Correspondence with microarray, normalized with 18S rRNA	Correlation (GAPDH)	Correlation (18S rRNA)	Significant difference between two groups (GAPDH)	Significant difference between two groups (18S rRNA)
1	TNFSF14	F	CTGTTGGTCAGCCAGCAGT	89	65℃	O(6.11)	O(2.36)				
		R	GAAAGCCCCGAAGTAAGACC	90							0.0065
2	MMP2	F	CAAGGACCGGTTCATTTGGC	91	60℃	O(3.82)	O(2.09)				
		R	GAACACAGCCTTCTCCTCCT	92							
3	SAA2	F	TGCTCGGGGGAACTATGATG	93	60℃	O(5.20)	O(2.47)				
		R	GGCCTGTGAGTCTCTGGATA	94							
4	COL1A1	F	GGAAGAGTGGAGAGTACTGG	95	60℃	O(2.56)	×(1.33)				
		R	ATCCATCGGTCATGCTCTCG	96							
5	COL1A2	F	GTATTCCTGGCCCTGTTGGT	97	60℃	O(2.92)	O(1.52)				
		R	CTCACCCTTGTTACCGCTCT	98							
6	DPYSL3	F	CTTTGAAGGGATGGAGCTGC	99	65℃	O(1.52)	×(0.78)				
		R	ATCGTACATGCCCCTTGGGA	100							
7	PPARD	F	GGCCTCTATCGTCAACAAGG	101	60℃	×(1.04)	××(0.40)				
		R	GCGTTGAACTTGACAGCAAA	102							
8	LUM	F	TACCAATGGTGCCTCCTGGA	103	60℃	×(1.38)	×(0.82)				
		R	CCACAGACTCTGTCAGGTTG	104							
9	MSTP032(RGS5)	F	CTGGAAAGGGCCAAGGAGAT	105	60℃	O(1.79)	×(1.03)				
		R	TCTGGGTCTTGGCTGGTTTC	106							
10	CRP	F	TGGCCAGACAGACATGTCGA	107	60℃	O(3.43)	O(1.60)				
		R	TCGAGGACAGTTCCGTGTAG	108							
11	TRIM38	F	TCTCTGGAGGCTGGAGAAAG	109	65℃	×(1.18)	××(0.49)				
		R	GTTTCCAGCTTCACAGCCCA	110							
12	S100A6	F	ATTGGCTCGAAGCTGCAGGA	111	60℃	O(1.83)	×(0.87)				
		R	GGAAGGTGACATACTCCTGG	112							
13	PZP	F	TACTCCAATGCAACCACCAA	113	65℃	O(4.39)	O(2.15)		r=0.717 (p=0.0171)		
		R	AACACAAGTTGGGATGCACA	114							
14	EMP1	F	TGGTGTGCTGGCTGTGCATT	115	60℃	O(1.65)	×(0.92)				
		R	GACCAGATAGAGAACGCCGA	116							
15	AI590053 (AL137672)	F	GTGAATGCCTCTGGAGTGGT	117	65℃	×(1.20)	××(0.46)				
		R	TTCTGTTCTGACGCCAAGTG	118							
16	MAP3K5	F	GTTCTAGCCAGTACTTCCGG	119	60℃	O(1.64)	×(0.69)			0.0528	
		R	ACTCGCTCCGAATTCTTGC	120							
17	TIMP1	F	ATTCCGACCTCGTCATCAGG	121	60℃	O(2.91)	O(1.62)				
		R	GCTGGTATAAGGTGGTCTGG	122							
18	GSTM1	F	GGACTTTCCCAATCTGCCCT	123	60℃	O(3.19)	O(1.64)				
		R	AGGTTGTGCTTGCGGGCAAT	124							
19	CSDA	F	AGGAGAGAAGGGTGCAGAAG	125	60℃	O(2.50)	×(1.09)				
		R	CCTTCCATAGTAGCCACGTC	126							
20	GSTM2	F	ACAACCTGTGCGGGGAATCA	127	65℃	O(1.82)	×(0.75)				
		R	GGTCATAGCAGAGTTTGGCC	128							
21	SGK	F	GCAGAAGGACAGGACAAAGC	129	60℃	O(1.75)	×(0.71)				
		R	CAGGCTCTTCGGTAAACTCG	130							
22	LMNA	F	ATGGAGATGATCCCTTGCTG	131	60℃	×(1.11)	××(0.50)				0.0282 (opposite)
		R	AGGTGTTCTGTGCCTTCCAC	132							0.0547 (opposite)
23	MGP	F	GCTCTAAGCCTGTCCACGAG	133	60℃	O(3.12)	O(1.83)				
		R	CGCTTCCTGAAGTAGCGATT	134							
24	LTBP2	F	GCGACACAGGAGTGTCAAGA	135	60℃	O(2.20)	×(1.21)				
		R	TGACCATGATGTAGCCCTGA	136							

With regard to "correspondence with microarray," the ratio of the late recurrence group and the early recurrence group was obtained from the results of quantitative PCR on 4 cases used for microarray analysis, and genes with the ratio of 1.5 or greater were indicated with O.
× indicates no difference, and ×× indicates an opposite correlation.
With regard to "correlation," genes exhibiting a correlation between the gene expression levels of 10 cases wherein the number of months of recurrence had been determined, and the period required for recurrence, were indicated with the r value and the p value.
In "significant difference between two groups," with regard to genes exhibiting a significant difference in expression levels between 6 cases of the recurrence within 24 months, and 8 cases of no recurrence for 48 months or more (the upper case) or 6 cases of no recurrence for 60 months or more (the lower case), p values (Mann-Whitney U test) were indicated.

As a result, it was found that when GAPDH was used as an internal standard gene, 19 out of the 24 gene candidates exhibiting up-regulation in the late recurrence group corresponded with the microarray results, and that among such genes, no genes exhibited a correlation with the recurrence period. In addition, when 18S rRNA was used as an internal standard gene, 9 out of the above 24 gene candidates corresponded with the microarray results, and among them, only 1 gene (PZP gene) exhibited a correlation with the recurrence period

A significant difference test was carried out on two groups, the late recurrence group and the early recurrence group. As a result, it was found that when GAPDH was

used as a standard gene, only one gene (MAP3K5 gene) exhibited a significant difference, and that when 18S rRNA was used as a standard gene, only one gene (TNFSF14 gene) exhibited a significant difference. On the contrary, there was one gene (LMNA gene), which had a significant difference, oppositely correlating to the recurrence period. Accordingly, this gene was identified as a gene up-regulated in nontumor tissues in the early recurrence group.

Subsequently, the results obtained by analyzing the 47 gene candidates (BNbad) up-regulated in nontumor tissues in the early recurrence group of type B hepatocellular carcinoma cases are shown in Table 15. Table 15 shows the analysis results obtained by quantitative PCR, which was performed on the cases shown in Table 13 as targets, under the conditions shown in Table 15 using GAPDH or 18S rRNA as an internal standard gene.

Table 15 Results of quantitative PCR of "genes up-regulated in nontumor tissues in early recurrence group of hepatitis B cases"

No.	Gene	Forward/ reverse	Primer sequence (5'-3')	SEQ ID NO.	Annealing temperature	Correspondence with microarray, normalized with GAPDH	Correspondence with microarray, normalized with 18S rRNA	Correlation (GAPDH)	Correlation (18S rRNA)	Significant difference between two groups (GAPDH)	Significant difference between two groups (18S rRNA)
1	CTH	F	TGAATGGCCACAGTGATGTT	137	60℃	○(4.47)	○(13.25)				
		R	CCATTCCGTTTTTGAAATGC	138							
2	OAT	F	TCGTAAGTGGGGCTATACCG	139	60℃	○(2.70)	○(11.89)				
		R	CTGGTTGGGTCTGTGGAACT	140							
3	PRODH	F	CTGACCACCGGGTGTACTTT	141	60℃	○(4.61)	○(22.30)				
		R	GACAAGTAGGGCAGCACCTC	142							
4	CYP3A7	F	GGAACCCGTACACATGGACT	143	60℃	× × (0.39)	× (1.27)				
		R	AACGTCCAATAGCCCTTACG	144							
5	DDT	F	CGCCCACTTCTTTGAGTTTC	145	60℃	× (1.04)	○(4.42)				
		R	CATGACCGTCCCTATCTTGC	146							
6	PGRMC1	F	TATGGGGTCTTTGCTGGAAG	147	65℃	× (1.15)	○(3.48)				
		R	GCCCACGTGATGATACTTGA	148							
7	AKR1C1	F	GGTCACTTCATGCCTGTCCT	149	60℃	× (1.32)	○(3.95)				
		R	TATGGCGGAAGCCAGCTTCA	150							
8	HGD	F	CACAAGCCCTTTGAATCCAT	151	60℃	○(1.61)	○(5.80)				
		R	TGTCTCCAGCTCCACACAAG	152							
9	FHR4	F	TTGAGAATTCCAGAGCCAAGA	153	60℃	× (0.83)	○(1.85)				
		R	CACCCATCTTCACCACACAC	154							
10	FST	F	AAGACCGAACTGAGCAAGGA	155	65℃	○(3.58)	○(6.80)				
		R	TTTTTCCCAGGTCCACAGTC	156							
11	COX4	F	-		-	-	-				
		R	-								
12	APP	F	CGGGCAAGACTTTTCTTTGA	157	60℃	× (1.28)	○(4.13)				
		R	TGCCTTCCTCATCCCCTTAT	158							
13	PSPHL	F	TCCAAGGATGATCTCCCACT	159	60℃	○(4.97)	○(5.44)				
		R	AGCATCCGATTCCTTCTTCA	160							
14	CYP1A1	F	TGATAAGCACGTTGCAGGAG	161	65℃	○(2.77)	○(11.30)				0.0389
		R	AAGTCAGCTGGGTTTCCAGA	162							0.0547
15	ZNF216	F	GGTGTCAGAGCCAGTTGTCA	163	60℃	○(1.84)	○(5.39)				
		R	AAATTTCCACATCGGCAGTC	164							
16	LEPR	F	CCACCATTGGTACCATTTCC	165	60℃	○(5.78)	○(14.99)				
		R	CCCCTCACCTGAACCTCATA	166							
17	TOM1L1	F	TTTTCTGGAACATTCAAATTCA	167	60℃	× (0.89)	○(2.61)				
		R	CACTTTTTGTCATCGCTGGA	168							
18	PECR	F	TGCAGTGGAATACGGATCAA	169	60℃	× (1.19)	○(3.49)				
		R	GGAAGCAGACCACAGAGGAG	170							
19	ALDH7A1	F	AGTGGAAGGTGTGGGTGAAG	171	65℃	× (1.34)	○(3.45)				
		R	CAACCATACACTGCCACAGG	172							
20	GNMT	F	CACTTAAGGAGCGCTGGAAC	173	60℃	○(1.82)	○(6.15)				
		R	TTTGCAGTCTGGCAAGTGAG	174							
21	OATPC	F	GCCACTTCTGCTTCTGTGTTT	175	60℃	× (1.27)	○(3.50)				
		R	TCCACCATAAAAGATGTGGAAA	176							
22	AKR1B10	F	CCTCCACTCATGTCCCATTT	177	60℃	○(2.92)	○(8.05)				
		R	TCAAGCCATGCTTTTCTGTG	178							
23	ANGPTL3	F	ATTTTAGCCAATGGCCTCCT	179	60℃	× (1.18)	○(3.37)				
		R	CACTGGTTTGCAGCGATAGA	180							
24	AASS	F	ATTGGTGAATTGGGATTGGA	181	60℃	○(2.04)	○(6.83)				
		R	GAAGCCCACCACAGTAGGAA	182							
25	CALR	F	TGGATCGAATCCAAACACAA	183	60℃	× (1.12)	○(2.77)				
		R	CTGGCTTGTCTGCAAACCTT	184							
26	BAAT	F	CTCCATCATCCACCCACTTT	185	60℃	× (1.15)	○(4.06)				
		R	GGAAGGCCAGCAAGTGTAGA	186							
27	PMM1	F	GCCAGAAAATTGACCCTGAG	187	60℃	× (1.04)	○(3.53)				
		R	CAGCTGCTCAGCGATCTTAC	188							
28	RABR	F	CCCTCATCGTGTCAAGTCAA	189	60℃	× (1.15)	○(3.78)				
		R	AGCATCAAACAGACCCAACC	190							
29	GLUL	F	TTGTTTGGCTGGGATAGAGG	191	60℃	× (0.85)	○(2.41)				
		R	GCTCTGTCCGGATAGCTACG	192							
30	CSHMT	F	CCCTACAAGGTGAACCCAGA	193	60℃	× (1.20)	○(3.33)				
		R	GGAGTAGCAGCTGGTTCCTG	194							

(Table 15, continued)

No.	Gene	Forward/ reverse	Primer sequence (5'-3')	SEQ ID NO.	Annealing temperature	Correspondence with microarray, normalized with GAPDH	Correspondence with microarray, normalize with 18S rRNA	Correlation (GAPDH)	Correlation (18S rRNA)	Significant difference between two groups (GAPDH)	Significant difference between two groups (18S rRNA)
31	UGT1A3	F	TGACAACCTATGCCATTTCG	195	60℃	×(0.89)	O(3.10)				
		R	CCACACAAGACCTATGATAGA	196							
32	HSPG1	F	CTCAAGGATGACGTGGGTTT	197	60℃	×(1.45)	O(4.17)				
		R	GATTTCCTCTGGCCAATTCA	198							
33	QPRT	F	AACTACGCAGCCTTGGTCAG	199	60℃	×(1.24)	O(3.91)				
		R	TGGCAGTTGAGTTGGGTAAA	200							
34	DEPP	F	GATGTTACCAATCCCGTTCG	201	60℃	O(2.68)	O(6.92)				
		R	TGGGCTCCTATATGCGGTTA	202							
35	CA2	F	TGCTTTCAACGTGGAGTTTG	203	65℃	O(1.73)	O(4.89)				
		R	CCCCATATTTGGTGTTCCAG	204							
36	FTHFD	F	CAAAATGCTGCTGGTGAAGA	205	60℃	×(1.28)	O(4.65)				
		R	GCCTCTGTCAGCTCAAGGAC	206							
37	LAMP1	F	GTCGTCAGCAGCCATGTTTA	207	60℃	××(0.61)	O(1.97)				
		R	GGCAGGTCAAAGGTCATGTT	208							
38	FKBP1A	F	GGGATGCTTGAAGATGGAAA	209	60℃	×(0.79)	O(1.79)				
		R	CAGTGGCACCATAGGCATAA	210							
39	BNIP3	F	GCTCCTGGGTAGAACTGCAC	211	60℃	×(1.00)	O(2.70)				
		R	GCCCTGTTGGTATCTTGTGG	212							
40	MAP3K12	F	TTGAGGAAATCCTGGACCTG	213	60℃	××(0.59)	O(1.52)				
		R	TTGAGGTCTCGCACCTTCTT	214							
41	ASS	F	CTGATGGAGTACGCAAAGCA	215	60℃	O(2.81)	O(9.16)				
		R	CTCGAGAATGTCAGGGGTGT	216							
42	ACTB	F	ACAGAGCCTCGCCTTTGC	217	60℃	×(0.74)	O(2.04)				
		R	CACGATGGAGGGGAAGAC	218							
43	PLAB	F	GAGCTGGGAAGATTCGAACA	219	60℃	O(2.57)	O(5.03)				
		R	AGAGATACGCAGGTGCAGGT	220							
44	ENO1L1	F	GAGATCTCGCCGGCTTTAC	221	60℃	×(0.75)	O(2.14)				
		R	CGCGAGAGTCAAAGATCTCC	222							
45	IGFBP3	F	CAGCTCCAGGAAATGCTAGTG	223	60℃	×(0.86)	O(2.81)			0.0528(逆)	
		R	GGTGGAACTTGGGATCAGAC	224							
46	UK114	F	GAGGGAAGGCTTAGCCATGT	225	60℃	×(1.11)	O(3.13)				
		R	TTGAAGGGTCCATGCCTATC	226							
47	ERF1	F	GCCTGTAAGTACGGGGACAA	227	60℃	×(1.16)	O(2.82)				
		R	CTCTTCAGCGTTGTGGATGA	228							

Although Gene Nos. 22 and 33 are genes common with CNbad, different sequences were used as PCR primers for Gene No. 22.
PCR was carried out on Gene No. 11 using 2 primer sets. However, since stable amplification did not achieved in any case, it was pending.
With regard to "correspondence with microarray," the ratio of the early recurrence group and the late recurrence group was obtained from the results of quantitative PCR on 4 cases used for microarray analysis, and genes with the ratio of 1.5 or greater were indicated with O.
× indicates no difference, and ×× indicates an opposite correlation.
There were no genes, which exhibited a correlation between the gene expression levels of 10 cases, wherein the number of months of recurrence had been determined, and the period required for recurrence.
In "significant difference between two groups," with regard to genes exhibiting a significant difference in expression levels between 6 cases of the recurrence within 24 months, and 8 cases of no recurrence for 48 months or more (the upper case) or 6 cases of no recurrence for 60 months or more (the lower case), p values (Mann-Whitney U test) were indicated.

As a result, it was found that when GAPDH was used as an internal standard gene, 16 gene corresponded with the microarray results, but that no genes significantly exhibited a correlation with the recurrence period. However, the IGFBP3 gene significantly exhibited an opposite correlation in the significant difference test between two groups. Accordingly, this gene was identified as a gene up-regulated in nontumor tissues in the late recurrence group.

In addition, when 18S rRNA was used as an internal standard gene, 45 genes corresponded with the microarray results, but that no genes significantly exhibited a correlation with the recurrence period. However, the CYP1A1 gene significantly exhibited a correlation in a significant difference test between two groups. Accordingly, this gene was identified as a gene up-regulated in nontumor tissues in the early recurrence group.

As stated above, the following 6 genes were identified as genes expressed in nontumor tissues, which can be used for prediction of the recurrence of cancer in type B hepatocellular carcinoma cases: the PZP gene, the MAP3K5 gene, the TNFSF14 gene, the LMNA gene, the CYP1A1 gene, and the IGFBP3 gene. The meanings of the aforementioned genes are as follows:

PZP gene: A pregnancy-zone protein gene

MAP3K5 gene: A mitogen-activated protein kinase kinase kinase 5 gene

TNFSF14 gene: A tumor necrosis factor (ligand) superfamily, member 14 gene

LMNA gene: A lamin A/C gene

CYP1A1 gene: A cytochrome P450, family 1, subfamily A, polypeptide 1 gene

IGFBP3 gene: An insulin-like growth factor binding protein 3 gene

Example 4

Selection of combination of genes used for distinguishing early recurrence group from late recurrence group

By combining several genes expressed in nontumor tissues used for prediction of the recurrence of type C or B hepatocellular carcinoma, which were obtained from the results of Examples 2 and 3, it becomes possible to carry out recurrence prediction more precisely. As such gene sets, many types of sets are conceived. Examples of the aforementioned combination are shown in Table 16.

Table 16 Examples of combinations of genes used for distinguishing hepatocellular carcinoma early recurrence group from late recurrence

Causal cancer	Early group	Late group	Normalization with GAPDH	Normalization with 18S rRNA
Type C hepatocellular cancer	< 24 months	> 40 months	VNN1 MRPL24	VNN1 CXCL9 GBP1 RALGDS
	Classification rate		88%	100%
Type B hepatocellular cancer	< 24 months	> 48 months	PRODH LMNA MAP3K12	LMNA LTBP2 COL1A2 PZP
	Classification rate		100%	100%

(1) Prediction of type C hepatocellular carcinoma

When GAPDH is used as an internal standard gene for normalization of gene expression in the distinction of an early recurrence group wherein the cancer has recurred within 24 months from a late recurrence group wherein the cancer has not recurred for 40 months or more, the gene expression level of VNN1 and that of MRPL24 may be examined. Otherwise, when 18S rRNA is used as an internal standard gene for normalization in the above distinction, the expression level of each gene of a gene set consisting of VNN1, CXCL9, GBP1, and RALGDS may be examined. The expression level of each of the aforementioned genes is assigned to a discriminant using a discriminant function coefficient obtained regarding each gene, and the obtained value is used for distinction. The expression level of the above gene group is analyzed. In the case of GAPDH normalization, the classification rate between the early recurrence group and the late recurrence group is found to be 88%, and in the case of 18S rRNA, the classification rate is found to be 100%.

(2) Prediction of type B hepatocellular carcinoma

When GAPDH is used as an internal standard gene for normalization in the distinction of an early recurrence group wherein the cancer has recurred within 24

months from a late recurrence group wherein the cancer has not recurred for 48 months or more, the expression level of each gene of a gene set consisting of PRODH, LMNA, and MAP3K12 may be examined. Otherwise, when 18S rRNA is used as an internal standard gene for normalization in the above distinction, the expression level of each gene of a gene set consisting of LMNA, LTBP2, COL1A2, and PZP may be examined. As described above, such expression levels are assigned to a discriminant, and the obtained values are used for distinction. The expression level of the above gene group is analyzed. In both cases of correlation with GAPDH and 18S rRNA, the classification rate between the early recurrence group and the late recurrence group is found to be 100%.

The meanings of the aforementioned genes are as follows:

PRODH gene: A proline dehydrogenase (oxidase) 1 gene

LTBP2 gene: A latent transforming growth factor beta binding protein 2 gene

COL1A2 gene: A collagen, type I, alpha 1 gene

MAP3K12 gene: A mitogen-activated protein kinase kinase kinase 12 gene

INDUSTRIAL APPLICABILITY

By identifying common genes derived from a patient and a healthy subject and cause-specific genes, it becomes possible to predict prognosis and recurrence. Accordingly, the thus identified genes can be used for diagnosis, the development of treatment methods, and a strategy of selecting a therapeutic agent (Taylor-made medicine).

Sequence Listing Free Text

SEQ ID NOS: 1 to 228: synthetic DNA